**Eurasian Chemical Communications** -

Original Research Article

http://echemcom.com

# Simultaneous determination of levodopa and tryptophan using a modified glassy carbon electrode

Mohammad Reza Aflatoonian<sup>a,b</sup>, Somayeh Tajik<sup>a,c,\*</sup>, Behnaz Aflatoonian<sup>c</sup>, Mehri-Saddat Ekrami-Kakhki<sup>d</sup>, Kouros Divsalar<sup>a</sup>, Iran Sheikh Shoaie<sup>e</sup>, Zahra Dourandish<sup>e</sup>, Mahdieh Sheikhshoaie<sup>f</sup>

fDepartment of Mining Engineering, Shahid Bahonar University of Kerman, Kerman, Iran

Received: 18 June 2019, Accepted: 04 December 2019, Published: 15 December 2019

#### **Abstract**

This work discusses the development of a new sensor for simultaneous determination of levodopa and tryptophan. The proposed sensor was a glassy carbon electrode modified with  $V_2O_5$  nanoparticles. Based on the excellent electrochemical properties of the modified electrode, a sensitive voltammetric method was used for individual and simultaneous determination of the levodopa and tryptophan in the aqueous solutions. The proposed method under the optimized conditions was utilized to determine the levodopa with linear range of  $0.06\text{--}250.0~\mu\text{M}$  and detection limit of  $0.02~\mu\text{M}$  (S/N=3). Finally, the applicability of the proposed sensor was verified by evaluation of levodopa and tryptophan in real sample.

**Keywords:** Levodopa; tryptophan; V<sub>2</sub>O<sub>5</sub> nanoparticles; glassy carbon electrode.

#### Introduction

Levodopa ((-)-3-(3,4-dihydroxyphenyl)-l-alanine, L-dopa) is an important chemical substance and a naturally occurring dietary supplement and psychoactive drug found in certain kinds of food and herbs and is synthesized from the essential amino acids l-phenylalanine and l-tyrosine in the mammalian body and brain [1]. Levodopa medication has been used for the most commonly prescribes for

Parkinson's disease. People with Parkinson's disease have depleted levels of dopamine, causing tremor, muscle stiffness or rigidity, slowness of movement (bradykinesia) and loss of balance. Since Dopamine cannot be administered directly as it does not cross the blood brain barrier readily, while after oral administration. levodopa is absorbed and converted dopamine by decarboxylase. Therefore, levodopa can alleviate the

\*Corresponding author: Somayeh Tajik Tel: +98 (913) 1965532, Fax: N/A

E-mail: tajik\_s1365@yahoo.com

<sup>&</sup>lt;sup>a</sup>Neuroscience Research Center, Kerman University of Medical Sciences, Kerman, Iran

<sup>&</sup>lt;sup>b</sup>Leishmaniasis Research Center, Kerman University of Medical Sciences, Kerman, Iran

<sup>&</sup>lt;sup>c</sup>Research Center for Tropical and Infectious Diseases, Kerman University of Medical Sciences, Kerman, Iran

dEsfarayen University of Technology, Esfarayen, Iran

<sup>&</sup>lt;sup>e</sup>Department of Chemistry, Faculty of Science, Shahid Bahonar University of Kerman, Kerman 76175-133, Iran

symptoms of Parkinson's disease, decreasing the muscular rigidity and tremors [1].

Tryptophan (TRP, 2-amino-3-(1Hindol-3-yl)-propionic acid), a vital constituent of proteins and a precursor for biologically important molecules such as the neurotransmitter serotonin, dopamine and the neurohormone melatonin. Moreover. it is indispensable component in human nutrition for building and keeping a positive nitrogen balance. Since TRP is not synthesized by the human organism and is not found in significant amounts in vegetables, it may be administered in the form of medications and food supplements [2]. TRP is one of the main components of drugs used to treat disorders such as depression and hypertension. Nonetheless, its improper metabolization has been suggested to be a possible cause of schizophrenia, hallucinations, and delusions, due to the formation of a toxic metabolite in the brain. Furthermore, high dosages of TRP can cause nausea, dizziness, and loss of appetite [3].

According to the above mentioned content and since supplementation may be beneficial for patients receiving levodopa, competition between tryptophan and levodopa may result in tryptophan malabsorption, leading to depression and other side effects of levodopa treatment. So, it is necessary and important for simultaneous determination of the levodopa and tryptophan in pharmaceutical biological samples.

A variety of analytical methods such as spectrophotometry [4,5], capillary electrophoresis [6,7], and chromatography methods [8,9], have been developed to measure the levodopa and tryptophan levels in different sample matrices. However, these methods are sometimes unsuitable

for routine analysis due to the requirement of expensive instruments, analysis time, and long sample pretreatment. Moreover, due to the electroactive properties of levodopa and tryptophan, those can be determined by electrochemical methods. Electrochemical technique is a good aforementioned alternative to the methods due to the faster response. higher selectivity, higher sensitivity, reproducibility, easy operation and lower cost [10-14].

Glassy carbon electrodes (GCEs) are very versatile as electrode material for trace level determination of organic as they provide high molecules sensitivity, negligible porosity, and good mechanical rigidity. Bare glassy carbon electrodes have always suffered serious problems such interference and electrode fouling, which can result in poor selectivity and reproducibility [15-17]. To resolve these problems, chemically modified electrodes (CME) with various materials have emerged as efficient and approach and versatile attracted considerable interest over the past two decades due to the advantages in terms of reduced costs, automatic and fast analysis, high sensitivity and selectivity Γ18-211.

The unprecedented achievements in nanoscience and nanotechnology in the recent era has strongly urged integrate researchers to the nanomaterial in electrode fabrication to achieve an ultrasensitive surface with improved selectivity. The reasons for their growing applications relates to their unique physical and chemical properties which entirely different from their bulk counterparts. Through different synthetic routes, it is now possible to develop nano-scaled materials with highly controlled shape, surface area, size, charge and physicochemical properties [23,24].

Vanadium oxide  $(V_2O_5)$  is the most oxide with semiconductor stable property and the energy gap of ~2.2 eV at room temperature. V<sub>2</sub>O<sub>5</sub> is of instance interest due to its layered structure, various phases, catalytic, good electrical and optical properties. Layered structure special characteristics (chemical sensing and electrochemical properties) of V<sub>2</sub>O<sub>5</sub> has attracted much interest. The good catalytic activity is the result of easy reduction and oxidation between the multiple oxidation states of vanadium in the V<sub>2</sub>O<sub>5</sub>. It was recently reported that  $V_2O_5$ possess electrocatalytic activity, where V<sup>5+</sup> ions play the dominant role for the electrooxidation reaction [25-27]. In the light of the above information, this work aims at developing a modified electrode for the sensitive determination of levodopa tryptophan, using V<sub>2</sub>O<sub>5</sub> nanoparticles to enhance the properties of glassy carbon electrodes. The resulting electrode is successfully applied for the determination of levodopa and tryptophan in the real samples.

## **Experimental**

Apparatus and Chemicals

An Autolab potentiostat/galvanostat (PGSTAT 302N, Eco Chemie, the Netherlands) was applied to measure the electrochemicals. General Purpose Electrochemical System (GPES) software was employed to control the experiments. conditions of conventional three-electrode cell was used at 25 ± 1 °C. An Ag/AgCl/KCl (3.0 M) electrode (Azar Electrode, Urmia, Iran), a platinum wire (Azar Electrode, Urmia, Iran), and the V<sub>2</sub>O<sub>5</sub>/GCE were used as the reference, auxiliary and working electrodes, respectively. pH was measured by a Metrohm 710 pH meter. Levodopa, tryptophan, and all the remaining reagents had an analytical grade. They have been prepared via Merck (Darmstadt, Germany). Orthophosphoric acid and the related salts that were above the pH range of 2.0-9.0 were used for preparing the buffer solutions. V<sub>2</sub>O<sub>5</sub> nanoparticles were synthesized in our laboratory as reported previously [27].

## **Preparing electrode**

 $V_2O_5$  nanoparticles have been used to coat the bare glassy carbon electrode. A stock solution of  $V_2O_5$ /GCE in 1 mL of aqueous solution has been prepared by distributing 1 mg of  $V_2O_5$  nanoparticles via ultra-sonication for 30 min, whereas 5  $\mu$ L of aliquots of the  $V_2O_5$  nanoparticles suspension solution has been cast on carbon working electrodes. Then, we waited until the solvent evaporation at room temperature.

## Preparing real samples

Samples of urine have been kept in a refrigerator directly after gathering. Ten millilitres of samples have been centrifuged for fifteen minutes at 2.000 rpm. The supernatant has been filtered by a 0.45 µm filter. Next, various volumes of solution has been transported into 25 a millilitres volumetric flask and diluted to the mark with PBS (pH= 7.0). This diluted urine samples were anaesthetized different amounts of levodopa and tryptophan. Content of levodopa and tryptophan have been analyzed using the suggested procedure by employing the standard addition method.

#### **Result and discussion**

Characterization of V<sub>2</sub>O<sub>5</sub>/NPs

The morphology of V<sub>2</sub>O<sub>5</sub>/NPs was assessed using scanning electron microscope (SEM). Figure 1 reveals the typical SEM images of the V<sub>2</sub>O<sub>5</sub>/NPs sample. It was found that the

synthesized V<sub>2</sub>O<sub>5</sub>/NPs have a spherical

shape with diameter ~ 35 nm.

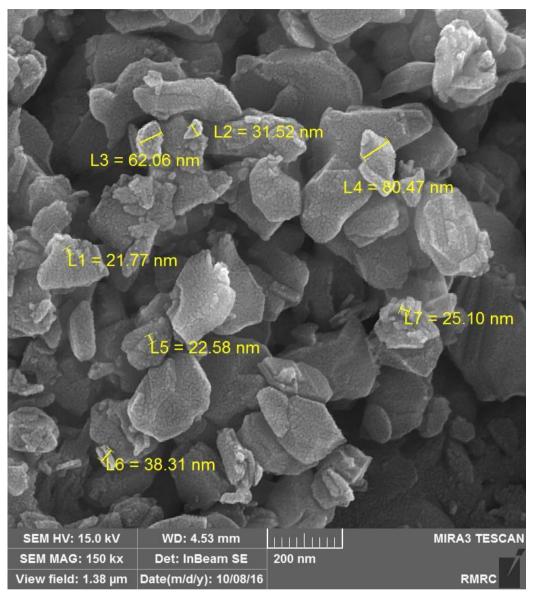


Figure 1. SEM image of V<sub>2</sub>O<sub>5</sub> nanoparticles

## Electrochemical properties of levodopa on V<sub>2</sub>O<sub>5</sub>/GCE surface

The electrochemical behaviour of levodopa is dependent on the pH value of the aqueous solution (Figure 2). Therefore, pH optimization of the solution seems to be necessary in order to obtain the electrocatalytic oxidation of levodopa. Thus the electrochemical behaviour of levodopa was studied in 0.1 M PBS in different pH values (2.0<

pH<9.0) at the surface of  $V_2O_5/GCE$  by CV. It was found that the electrocatalytic oxidation of levodopa at the surface of  $V_2O_5/GCE$  was more favoured under neutral conditions than in acidic or basic medium, because the obtained current was more than other pHs. Thus, the pH 7.0 was chosen as the optimum pH for electrocatalysis of levodopa oxidation at the surface of  $V_2O_5/GCE$ .

HO OH 
$$-2H^+$$
  $-2e^-$  OH NH<sub>2</sub>

Figure 2. Electrochemical oxidation of levodopa

Figure 3 shows responses of CV to electro-oxidation of  $100.0~\mu M$  levodopa at the unmodified GCE (curve b) and  $V_2O_5/GCE$  (curve a). The peak potential occurs at 350 mV due to levodopa oxidation, which is around 300 mV more negative than the unchanged GCE. Furthermore,

V<sub>2</sub>O<sub>5</sub>/GCE exhibits very high anodic peak currents for levodopa oxidation than that of the unchanged GCE. This showed a significant improvement of the electrode performance toward levodopa oxidation by changing the constant GCE with V<sub>2</sub>O<sub>5</sub> nanoparticles.

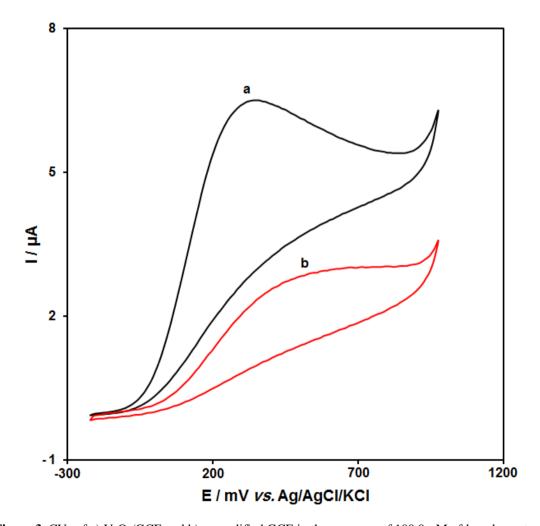
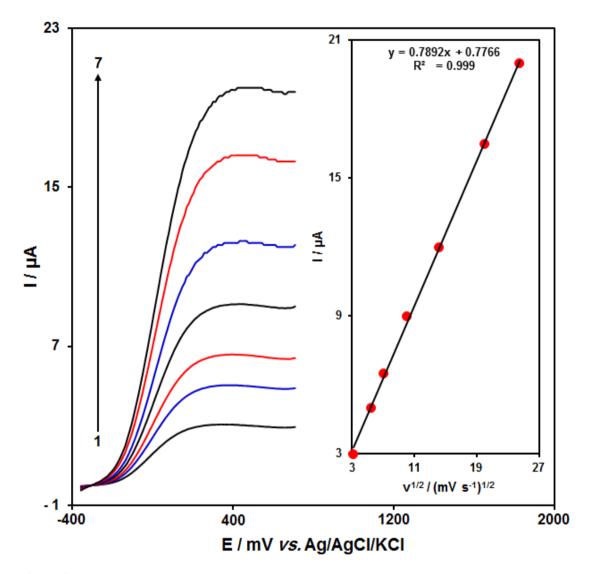


Figure 3. CVs of a)  $V_2O_5/GCE$  and b) unmodified GCE in the presence of 100.0  $\mu M$  of levodopa at pH 7.0. In all cases, the scan rate was 50 mV s<sup>-1</sup>

#### Effect of scan rate

Researchers investigated the impact of the rates of potential scan on levodopa oxidation current (Figure 4). Findings indicated induction of enhancement in the current of the peak by the increased potential scan rate. Additionally, diffusion in oxidation processes are monitored, as inferred by the linear dependence of the anodic peak current (Ip) on the square root of the potential scan rate ( $v^{1/2}$ ) [28].



**Figure 4.** LSVs of  $V_2O_5$ /GCE in 0.1 M PBS (pH 7.0) containing 100.0  $\mu$ M of levodopa at various scan rates; numbers 1–7 correspond to 10, 30, 50, 100, 200, 400 and 600 mVs<sup>-1</sup>, respectively. Inset: Variation of anodic peak current  $\nu s$ . square root of scan rate

## Chronoamperometric analysis

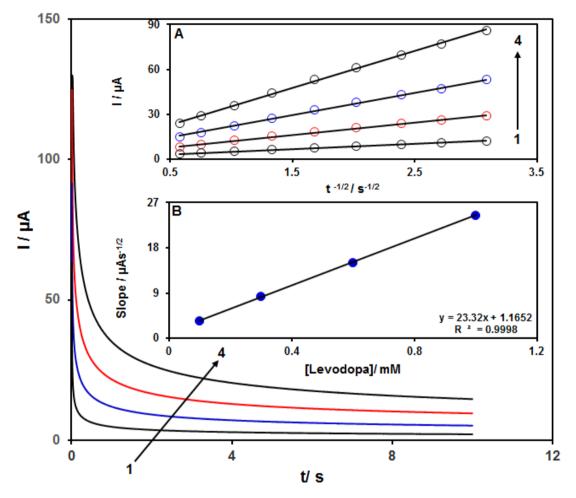
Chronoamperometric measurements of levodopa at V<sub>2</sub>O<sub>5</sub>/GCE were conducted by adjusting the working electrode potential at 0.4 V versus Ag/AgCl/KCl (3.0 M) for different concentrations of levodopa (Figure 5) in PBS (pH 7.0), respectively. For electroactive materials

(levodopa inthis case) with a diffusion coefficient of D, the Cottrell equation describes current seen for electrochemical reaction at the mass transport limited condition [28].

$$I = nFAD^{1/2}C_b\pi^{-1/2}t^{-1/2}$$

where D and  $C_b$  respectively represent diffusion coefficient (cm<sup>2</sup> s<sup>-1</sup>) and bulk concentration (mol cm<sup>-3</sup>). Experimental plots of I versus t<sup>-1/2</sup> were used with the best fits for various concentrations of levodopa (Figure 5A). Then, the

resultant straight lines slopes were drawn against levodopa concentrations (Figure 5B). According to the resultant slope and the Cottrell equation, mean value of D was  $4.6\times10^{-5}$  cm<sup>2</sup>/s for levodopa.



**Figure 5.** Chronoamperograms obtained at V<sub>2</sub>O<sub>5</sub>/GCE in 0.1 M PBS (pH 7.0) for different concentrations of levodopa. The numbers 1-4 correspond to 0.1, 0.3, 0.6 and 1.2 mM of levodopa. Insets: (A) Plots of I vs. t<sup>-1/2</sup> obtained from chronoamperograms 1–5. (B) Plot of the slope of the straight lines against levodopa concentrations

#### Calibration curve and LOD

The electro-oxidation peak currents of levodopa at V<sub>2</sub>O<sub>5</sub>/GCE surface can be applied to define levodopa in the solution. Since the increased sensitivity and more suitable properties for analytical utilizations are considered as the benefits of differential pulse voltammetry (DPV), V<sub>2</sub>O<sub>5</sub>/GCE in 0.1 M PBS consisting of different distinct concentrations of levodopa was used to

conduct DPV experiments (Figure 6). It was found that the electrocatalytic peak currents of levodopa oxidation at  $V_2O_5/GCE$  surface linearly depended on levodopa concentrations above the range of 0.06-200.0  $\mu M$  (with a correlation coefficient of 0.999), while determination limit (3 $\sigma$ ) was achieved to be 0.02  $\mu M$ . In the case of tryptophan electrocatalytic peak currents of tryptophan oxidation at the

surface of  $V_2O_5/GCE$  were linearly dependent on the tryptophan concentrations, over the range of  $5.0\times10^{-6}$  M $-4.0\times10^{-4}$  M (with a

correlation coefficient of 0.999) and the detection limit  $(3\sigma)$  was obtained  $2.0 \times 10^{-6}$  M.

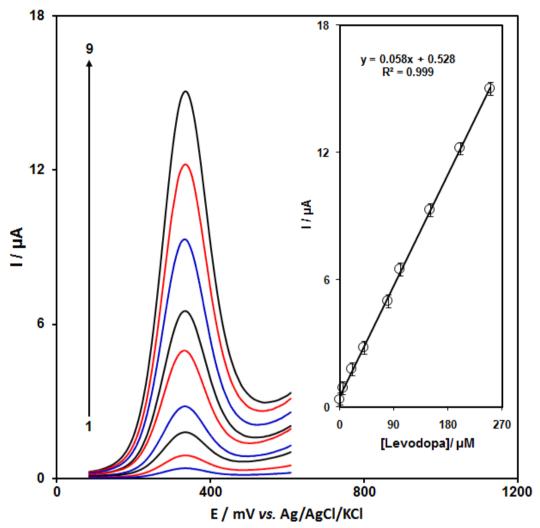
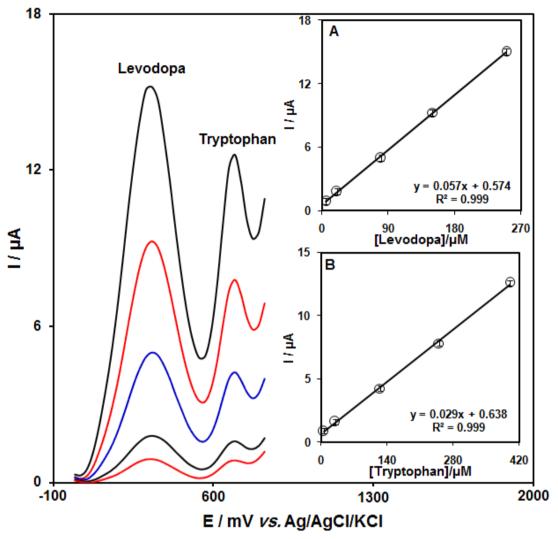


Figure 6. DPVs of  $V_2O_5/GCE$  in 0.1 M PBS (pH 7.0) containing different concentrations of levodopa. Numbers 1-9 correspond to, 0.06, 5.0, 20.0, 40.0, 80.0, 100.0, 150.0, 200.0 and 250.0  $\mu$ M of levodopa. The inset shows the plot of the peak current as a function of the levodopa concentration in the range of 0.06-250.0  $\mu$ M

## Simultaneous determination of levodopa and tryptophan

To the best of our knowledge, there is no report on using a GCE modified with  $V_2O_5$  nanoparticles for determining levodopa and tryptophan. Moreover, due to reality that electrochemical detection of levodopa in the front of tryptophan with the help of un-modified electrodes has the caveat of interventions by tryptophan because of relative adjacent oxidation

capacities of the two specimens, it can be regarded a crucial phase. Such a phase has been conducted simultaneous alterations of analytes concentrations and achieving DPVs (Figure 7). Findings reported certain anodic at 330 and 700 mV for levodopa and tryptophan oxidation, proving using the  $V_2O_5/GCE$ , these two analytes can be detected without severe interventions from each another.



**Figure 7.** DPVs achieved at  $V_2O_5/GCE$  surface in 0.1 M PBS (pH 7.0) consisting of various concentrations of levodopa and tryptophan. DPVs from internal to external respectively is corresponding to 5.0+5.0, 20.0+30.0, 80.0+125.0, 150.0+250.0 and 250.0+400.0  $\mu$ M of levodopa and tryptophan. Insets: (a) plot of Ip versus levodopa concentration and (B) plot of Ip versus tryptophan concentration

## **Analysis of real samples**

To assess the applicability of the modified electrode to determine the levodopa and tryptophan in real samples, the described method was applied to the determination of the levodopa and tryptophan in urine

sample. Therefore, the standard addition technique was applied (Table 1). Acceptable recoveries of levodopa and tryptophan were observed, and reproducible results were shown with regard to the mean relative standard deviation (R.S.D.).

Method	Electrode	Modifier	Linear	Detection	Ref
			range (µM)	limit (µM)	
Voltammetry	Glassy carbon	rGO	2.0-100.0	1.13	29
Voltammetry	Carbon paste	FCMCNPE	2.0-500.0	1.2	30
Voltammetry	Carbon ionic liquid	Nafion/Co(OH)2-MWCNTs	0.25-225.0	0.12	31
Voltammetry	Glassy carbon	poly(2,5-diamino benzene	1.0-200.0	0.41	32
		sulfonic acid)-β-cyclo dextrine			
Voltammetry	Carbon paste	BNH/TiO2	2.0-600.0	0.2	33
Voltammetry	Glassy carbon	$ m V_2O_5$	0.06-250	0.02	This
					work

Table 1. Comparison of the efficiency of some modified electrodes used in detection of levodopa

#### Conclusion

In this research study, a V<sub>2</sub>O<sub>5</sub>/GCE was developed for the electrochemical simultaneous determination levodopa and tryptophan. In comparison with the previously reported methods for electrochemical determination of levodopa, the method presented has many advantages, such as simple fabrication, low detection limit and low cost. The V<sub>2</sub>O<sub>5</sub>/GCE shows a good electrochemical response for determination of levodopa with the wide linear range between 0.06 and 250.0 µM and the limit of detection was calculated to be 0.02 µM. Moreover, V<sub>2</sub>O<sub>5</sub>/GCE operated well in the simultaneous determination of levodopa and tryptophan real samples with great accuracy and precision.

## Acknowledgements

The authors acknowledge the financial support provided for this project (Project No. 98000293 and ethics code EC/98-29/KNRC) by Neuroscience Research Center, Kerman University of Medical Sciences, Kerman, Iran.

#### References

- [1] P. Damier, E.C. Hirsch, Y. Agid, A.M. Graybiel, *Brain*, **1999**, *122*, 1437-1448.
- [2] B. Kaur, T. Pandiyan, B. Satpati, R. Srivastava, *Coll. Surf. B Biointerfaces*, **2013**, *111*, 97-106.

- [3] I.A. Mattioli, M. Baccarin, P. Cervini, É.T. Cavalheiro, *J. Electroanal. Chem.*, **2019**, 835, 212-219.
- [4] P. Nagaraja, K.S. Murthy, K.S. Rangappa, N.M. Gowda, *Talanta*, **1998**, *46*, 39-44.
- [5] I. Molnár-Perl, M. Pintér-Szakács, *Anal. Biochem.*, **1989**, *177*, 16-19.
- [6] W.W. He, X.W. Zhou, J.Q. Lu, *J. Chromatogr A*, **2006**, *1131*, 289-292.
- [7] K.D. Altria, P. Harkin, M.G. Hindson, *J. Chromatogr B: Biomed. Sci. Appl.*, **1996**, *686*, 103-110.
- [8] M. Doležalová, M. Tkaczykova, *J. pharmaceut. Biomed. Anal.*, **1999**, *19*, 555-567.
- [9] S.Y. Lee, K.M. Park, S.H. Jo, H.G. Nam, S. Mun, *J. Chromatogr A*, **2011**, *1218*, 1195-1202.
- [10] S. Mohammadi, A. Taheri, Z. Rezayati-Zad, *Prog. Chem. Biochem. Res.*, **2018**, *I*, 1-10.
- [11] H. Beitollahi, Z. Dourandish, S. Tajik, M.R. Ganjali, P. Norouzi, F. Faridbod, *J. Rare Earths*, **2018**, *36*, 750-757.
- [12] G. Vinodhkumar, R. Ramya, M. Vimalan, I. Potheher, A. Cyrac Peter, *Prog. Chem. Biochem. Res.*, **2018**, *1*, 40-49.
- [13] M.R. Ganjali, Z. Dourandish, H. Beitollahi, S. Tajik, L. Hajiaghababaei, B. Larijani, *Int. J. Electrochem. Sci*, **2018**, *13*, 2448-2461.

- [14] M. Eldefrawy, E.G.A. Gomaa, S. Salem, F. Abdel Razik, *Prog. Chem. Biochem. Res.*, **2018**, *1*, 11-18.
- [15] S.E. Baghbamidi, H. Beitollahi, S. Tajik, R. Hosseinzadeh, *Int. J. Electrochem. Sci.*, **2016**, *11*, 10874-10883.
- [16] Q. He, J. Liu, X. Liu, G. Li, D. Chen, P. Deng, J. Liang, *Nanomaterials*, **2018**, *8*, 194-208.
- [17] S. Tajik, H. Beitollahi, P. Biparva,
  J. Serb. Chem. Soc., 2018, 83, 863-874.
  [18] H. Qi, C. Zhang, Electroanalysis,
  2005, 17, 832-838.
- [19] Y. Wang, Y. Li, L. Tang, J. Lu, J. Li, *Electrochem. Commun*, **2009**, *11*, 889-892.
- [20] M. Mazloum-Ardakani, H. Beitollahi, M.K. Amini, F. Mirkhalaf, B.F. Mirjalili, A. Akbari, *Analyst*, **2011**, *136*, 1965-1970.
- [21] H. Jin, C. Zhao, R. Gui, X. Gao, Z. Wang, *Anal. Chim. Acta*, **2018**, *1025*, 154-162.
- [22] H.M. Moghaddam, H. Beitollahi, S. Tajik, M. Malakootian, H.K. Maleh, *Environ. Monit. Assess.*, **2014**, *186*, 7431-7441.
- [23] G.C. Han, X. Su, J. Hou, A. Ferranco, X.Z. Feng, R. Zeng, H.B. Kraatz, *Sens. Actuators B Chem.*, **2019**, 282, 130-136.
- [24] A. Taherkhani, T. Jamali, H. Hadadzadeh, H. Karimi-Maleh, H.

- Beitollahi, M. Taghavi, F. Karimi, *Ionics*, **2014**, *20*, 421-429.
- [25] R. Suresh, K. Giribabu, R. Manigandan, S.P. Kumar, S. Munusamy, S. Muthamizh, V. Narayanan, *Sens. Actuators B Chem.*, **2014**, *202*, 440-447.
- [26] D.P. Nair, T. Sakthivel, R. Nivea, J.S. Eshow, V. Gunasekaran, *J. Nanosci. Nanotechnol*, **2015**, *15*, 4392-4397.
- [27] M. Sheikhshoaie, I. Sheikhshoaie, M. Ranjbar, *J. Mol. Liq.*, **2017**, *231*, 597-601.
- [28] A.J. Bard, L.R. Faulkner, second ed., Wiley, New York, NY, **2001**.
- [29] S.Y. Yi, J.H. Lee, H.G. Hong, *J. Applied Electrochem.*, **2014**, *44*, 589-597.
- [30] H. Yaghoubian, H. Karimi-Maleh, M.A. Khalilzadeh, F. Karimi, *Int. J. Electrochem. Sci*, **2009**, *4*, 993-1003.
- [31] A. Babaei, A.R. Taheri, M. Aminikhah, *Electrochim. Acta*, **2013**, 90, 317-325.
- [32] M. Aslanoglu, A. Kutluay, S. Goktas, S. Karabulut, *J. Chem. Sci.*, **2009**, *121*, 209-215.
- [33] M. Mazloum-Ardakani, H. Beitollahi, M.A. Sheikh-Mohseni, H. Naeimi, *J. Iran. Chem. Soc.*, **2012**, *9*, 27-34.

**How to cite this manuscript:** Mohammad Reza Aflatoonian, Somayeh Tajik, Behnaz Aflatoonian, Mehri-Saddat Ekrami-Kakhki, Kouros Divsalar, Iran Sheikh Shoaie, Zahra Dourandish, Mahdieh Sheikhshoaie. Simultaneous determination of levodopa and tryptophan using a modified glassy carbon electrode. *Eurasian Chemical Communications*, 2020, 2(4), 505-515.